# TOTAL NITROGEN DETERMINATION IN AMMONIACAL, AMIDIC AND NITRATE NITROGEN CONTAINING FERTILIZERS BY THE RANEY CATALYST METHOD

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### ABSTRACT

This paper presents the optimized conditions for the determination of total nitrogen in ammoniacal, amidic, and nitrate nitrogen containing fertilizers by the Raney catalyst method using a Buchi K-436 speed digester and a Buchi K-360 distillation unit.

Keywords: Fertilizers, nitrogen, raney, ammonical, amidic, nitrate

## **INTRODUCTION**

Nitrogen is one of the nutrients that mostly limit plant growth. Therefore, fertilizers normally contain at least one form of nitrogen, mainly ammoniacal, amidic, or nitrate nitrogen, as exemplified in Table 1.

	Туре	Examples
N-NH <sub>4</sub> <sup>+</sup>	Ammoniacal	Monoammonium phosphate (NH <sub>4</sub> H <sub>2</sub> PO <sub>4</sub> ) (MAP)
		Ammonium sulfate (NH <sub>4</sub> SO <sub>4</sub> )
N-amidic	Amidic	Urea (CO(NH <sub>2</sub> ) <sub>2</sub> )
N-NO <sub>3</sub> <sup>-</sup>	Nitrate	Calcium nitrate (Ca(NO <sub>3</sub> ) <sub>2</sub> ) (CN)
N-NH4+/ N-	Ammoniacal +	Ammonium nitrate (NH4NO3) (AN)
NO3-	nitrate	Calcium ammonium nitrate
		(5Ca(NO3)2.NH4NO3.10H2O) (CAN)

 Table 1. Some types of nitrogen used in fertilizers

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In the soil, the amidic nitrogen (N-amidic) has to be hydrolyzed to ammoniacal nitrogen (N-NH4+), which then suffers nitrification by bacteria to the nitrate form (N-NO3-). Only nitrate is taken up by plants, rapidly and at high rates. Nitrate-based fertilizers have been considered the most efficient nitrogen sources, since they are immediately available as nutrients for plant uptake and cause less acidification of the soil than urea or ammonium-based fertilizers. However, granular nitrates absorb water from air and ammonium nitrate (AN) is a detonable substance, which has caused several disasters through the 20th century (BRABAUSKAS, 2016).

The analysis of total nitrogen is usually carried out by the Kjeldahl method (1883), which starts by the digestion of nitrogen-containing compounds in the presence of a catalyst (such as Hg, Cr, Ni-Al alloy, or Cu) and sulfuric acid, which is a weak oxidant (KIRK, 1950; SÁEZ-PLAZA et al., 2013). Potassium sulfate is used to raise boiling point (ABRAMS et al., 2014; CAMPBELL & HANNA, 1937; HILLER et al., 1948) and the rate of digestion (KIRK, 1950). The quantity of sulfuric acid has to exceed the amount of K<sub>2</sub>SO<sub>4</sub> to prevent ammonia losses, and the usual acid and salt contents (15 mL H<sub>2</sub>SO<sub>4</sub> and 15 g K<sub>2</sub>SO<sub>4</sub>, which is divided in two portions, 7.5 g K<sub>2</sub>SO<sub>4</sub> dissolved in the acid, and 7.5 g K<sub>2</sub>SO<sub>4</sub> added later as powder) produce a digestion temperature of about 380 °C, without reaching 400 °C, which may lead to pyrolityc loss of nitrogen (SÁEZ-PLAZA et al., 2013). The use of a high K<sub>2</sub>SO<sub>4</sub> to H<sub>2</sub>SO<sub>4</sub> ratio will cause loss of ammonia (BARKER & VOLK, 1964). After digestion, nitrogen is reduced to ammonia and presented in the form of ammonia, which is collected in an acidic solution. Finally, an acid-base titration determines the amount of ammonia and consequently the nitrogen content.

The Manual of Oficial Analytical Methods for Fertilizers and Correctives of the Brazilian Ministry of Agriculture, Livestock and Food Supply (MAPA, 2017a, b) addopts the Raney catalyst method (BRABSON & BURCH, 1964; BRABSON & WOODIS, 1969) for nitrogen determination, presented as macro- and micromethods. Among the Raney catalyst powders (alloys of aluminum with other metals), the alloy containing Ni and Al is the most efficient reductant of nitrates in acid medium (BRABSON & BURCH, 1964), usually in the proportion 50 % Ni : 50 % Al (JOHNSON & MILLER, 1974). Nascent hidrogen is generated by reaction of Al with the acid and reduces nitrate to ammonia (BRABSON & BURCH, 1964). The accurracy of the reaction increases when carried out in a solution containing sulfuric acid and part of the potassium sulphate (BRABSON & WOODIS, 1969). In the nitrate analysis using the Raney catalyst method, the

maximum amount 42 mg nitrate is set (BRABAUSKAS, 2016; BRABSON & BURCH, 1964; MAPA 2017a, b) in order to ensure that at least 1.5 g Raney alloy is available to reduce 42 mg nitrate (BRABSON & WOODIS, 1969). The reduction of nitrate to ammonia can also be carried out using salicilic acid (DICKINSON, 1954; RANKER, 1926; SAH, 1994) using for instance HgO or metallic Hg (AOAC 955.04-D method) or using CuSO<sub>4</sub>.5H<sub>2</sub>O (MAPA, 2017a, b), however the method fails in the presence of chloride and is not suitable for liquid fertilizers (BRABSON & BURCH, 1964).

In the LFDA/PA fertilizers laboratory, since 2019 the amount of nitrate fertilizers received for analysis has increased considerably. Usually the samples are sent for analysis without description of the fertilizer nitrogen type. Therefore, the analysis is initially carried out by the Raney macromethod and then, if necessary, it is repeated by the same method with reduced sample uptake or the analysis is performed by the Raney micromethod. This article describes the optimized conditions to carry out the Raney catalyst macro- and micromethods in ammonical, amidic, and nitrate containing fertilizers using a Buchi K-439 SpeedDigester and a Buchi K-360 distillation unit.

#### **MATERIALS AND METHODS**

#### **Instruments and apparatus**

Samples and standards were weighed in Sartorius Cubis MSA analytical balances (Sartorius, São Bernardo do Campo, Brazil). The digestion was carried out in a Buchi K-436 SpeedDigester (Buchi, Valinhos, Brazil), connected to a Buchi scrubber K-415, while the distillation was performed using a Buchi KjelFlex K-360 distillation unit. For the acid-base titration, calibrated digital Titrette burettes (Brand, Wertheim, Germany) or glass burettes were used.

#### **Reagents and chemicals**

Ultrapure water (resistivity 18.2 M $\Omega$  cm) was obtained from a Milli-Q Integral 15 purification system (Merck Millipore, São Paulo, Brazil) and used throughout. Potassium sulfate (Merck, Rio de Janeiro, Brazil), sulfuric acid (Merck Millipore, São Paulo, Brazil), Raney catalyst (50 % Ni and 50 % Al) (Dinâmica, Indaiatuba, Brazil), copper sulfate pentahydrate (Merck, São Paulo, Brazil), sodium thiosulfate (Sigma-Aldrich, São Paulo, Brazil), sodium hydroxide (Dinâmica, Indaiatuba, Brazil), boric acid (J. T. Baker, Phillipsburg, Brazil), methyl red (Sigma-

Aldrich, São Paulo, Brazil), bromocresol green (Neon, Suzano, Brazil), and potassium nitrate 99 % (Química Moderna, Barueri, Brazil) were chemical grade. Kjeldahl tablets were Merck (Merck Millipore, São Paulo, Brazil) and consisted of a mercury- and selenium-free catalyst containing 47.7% Na<sub>2</sub>SO<sub>4</sub>, 47.7% K<sub>2</sub>SO<sub>4</sub>, 2.8% TiO<sub>2</sub>, and 1.8% CuSO<sub>4</sub>. Ammonium sulphate, MAP, and urea were obtained as fertilizer raw materials, analyzed by the Raney catalyst micromethod, and validated in our laboratory for homogeneity and stability in order to be used as quality control samples. The nitrate fertilizer standard reference material was SRM NIST 193 (NIST, Gaithersburg, United States) and contained 13.84 % w/w of nitrogen. The K<sub>2</sub>SO<sub>4</sub>-H<sub>2</sub>SO<sub>4</sub> solution was prepared using 20 % V V<sup>-1</sup> H<sub>2</sub>SO<sub>4</sub> and 10.67 % w V<sup>-1</sup> K<sub>2</sub>SO<sub>4</sub>.

### **Fertilizers analysis**

The analysis of fertilizers was performed as summarized in Figure 1. The samples and reagents listed in Table 2 were added to 300-mL Buchi SpeedDigester sample tubes and the digestion was performed according to the conditions of Table 3. After the mixtures were cooled down, the distillation was carried out as shown in a Buchi K-360 distillation unit using the parameters of Table 4. Finally, the distilled solutions were titrated using 0.25 M sulfuric acid (macromethod) or 0.1 sulfuric acid (micromethod) using methyl red and bromocresol green as indicators.



**Figure 1.** Determination of total nitrogen in ammoniacal, amidic and nitrate nitrogen containing fertilizers by the Raney catalyst method using a Buchi K-436 SpeedDigester and a Buchi K-360 distillation unit.

		Raney macromethod		Raney micromethod
Fertilizer / nitrogen type	Granular, not nitrate	Granular, nitrate	Liquid, not nitrate	Granular and liquid, nitrate and non-nitrate
Sample mass	0.5 g sample	Equivalent to 21	2.5 g sample	1.0 g sample
		mg N-NO <sub>3</sub> - <sup>(1)</sup>		
Water			250 mL	250 mL
Aliquot			Equivalent to 10	Equivalent to 2.5 mg to
			mg to 40 mg N	15 mg N
Raney catalyst	0.85 g	0.85 g	0.85 g	0.70 g
H <sub>2</sub> SO <sub>4</sub> -	75 mL <sup>(2)</sup>	75 mL <sup>(2)</sup>	75 mL <sup>(2)</sup>	
K <sub>2</sub> SO <sub>4</sub> solution				
CuSO <sub>4</sub> .5H <sub>2</sub> O	0.5 g <sup>(3)</sup>	0.5 g <sup>(3)</sup>		
K <sub>2</sub> SO <sub>4</sub>	7.5 g <sup>(3)</sup>	7.5 g <sup>(3)</sup>	7.5 g <sup>(3)</sup>	
H <sub>2</sub> SO <sub>4</sub>				5 mL
Water				From the aliquot

# **Table 2.** Fertilizers preparation for digestion.

<sup>(1)</sup> In case of KNO<sub>3</sub> (13.72 % N) and NIST 193 (13.84 % N), sample uptake was 0.15 g (equivalent to 42 mg nitrate).

 $^{(2)}$  75 mL of K<sub>2</sub>SO<sub>4</sub>-H<sub>2</sub>SO<sub>4</sub> solution contains 15 mL H<sub>2</sub>SO<sub>4</sub> and 8.0025 g K<sub>2</sub>SO<sub>4</sub>, which corresponds to 20 % V V<sup>-1</sup> H<sub>2</sub>SO<sub>4</sub> and 10.67 % w V<sup>-1</sup> K<sub>2</sub>SO<sub>4</sub>.

<sup>(3)</sup> Alternatively, K<sub>2</sub>SO<sub>4</sub> and CuSO<sub>4</sub>.5H<sub>2</sub>O were replaced by one Merck Kjeldahl tablet.

### Table 3. Digestion conditions at the Buchi K-436 SpeedDigester.

Raney macromethod	Raney micromethod				
Preheat: 370 °C					
Step 1: 370 °C 1 min	Preheat: 370 °C				
Step 2: 350 °C 13 min <sup>(1)</sup>	Step 1: 370 °C 45 min				
Step 3: 370 °C 60 min	Step 2: 50 °C 0 min				
Step 4: 50 °C 0 min					

<sup>(1)</sup> 13 minutes corresponds to 3 minutes necessary to drop the temperature from 370 °C to 350 °C, followed by 10 minutes at 350 °C.

#### TOTAL NITROGEN DETERMINATION IN AMMONIACAL, AMIDIC AND NITRATE NITROGEN CONTAINING FERTILIZERS BY THE RANEY CATALYST METHOD

Reagents /	Use	Raney	Raney
Distillation parameters		macromethod	micromethod
Water	Added to sample tube	50 mL	50 mL
$Na_2S_2O_3.5H_2O \ 8\% \ w/V$	Added to sample tube	3 mL	-
NaOH 32% w/V	Added to sample tube	78 mL	35 mL
H <sub>3</sub> BO <sub>3</sub> 40% w/V	Added to collector flask	50 mL	5 mL
Water	Added to collector flask	-	40 mL
Reaction time	-	5 s	5 s
Steam power	-	100 %	100 %
Distillation time	-	300 s	300 s

#### Table 4. Distillation conditions for the Buchi KjelFlex K-360 distillation unit.

### **RESULTS AND DISCUSSION**

#### Sample preparation, digestion and distillation methods

The macro- and micromethods of the Raney catalyst used in the study are based on the Manual of Official Analytical Methods for Fertilizers and Correctives of the Brazilian Ministry of Agriculture, Livestock and Food Supply (MAPA, 2017a, b). However, due to the use of Buchi equipment (K-436 SpeedDigester and K-360 distillation unit), some adjustments were necessary, as summarized in Tables 2, 3 and 4 and discussed below.

#### Raney catalyst macromethod: digestion temperature

Regarding the digestion temperature, the Buchi conditions were first set for N-NO<sub>3</sub><sup>-</sup> containing fertilizers, whose digestion parameters are the most difficult to optimize. Once these conditions were set (Table 2), they were satisfactorily tested for the other nitrogen forms (ammoniacal and amidic). The MAPA manual describes the use of an initial digestion temperature that brings 100 mL water to boiling in 5 minutes, which should then be reduced to a temperature that brings 100 mL water to boiling in 10 minutes, and finally be raised again to the initial temperature, which shall be kept until  $H_2SO_4$  white fumes cease and the mixture becomes clear. In the K-439 Buchi SpeedDigester, the temperature of 350 °C brings water to boil in 10 minutes, however the temperature ramp developed by this digester is not quick enough to bring water to boil in 5 minutes, no matter how high the initial temperature of 370 °C (this step usually takes

11 minutes), which was kept for 1 minute (step 1), then the temperature was reduced to 350 °C and kept for 10 minutes (step 2, 13 minutes in total), and finally the temperature was raised again to 370 °C and maintained until the white fumes ceased (step 4, 60 minutes), which serves as an indication that the digestion of ammoniacal, nitrate, and urea nitrogen was completed (BRABSON & WOODIS, 1969; MAPA, 2017a, b).

A longer digestion method, in which the step 3 at 370 °C was increased from 60 to 90 minutes, was also tested, in order to extend the digestion by 30 minutes after the white fumes ceased, as recommended by Brabson & Woodis (1969), however no improvements in nitrogen recoveries were observed. Additionally, a two-step digestion method was also tested, as nitrogen digestion is usually carried out (addition of Raney catalyst, H<sub>2</sub>SO<sub>4</sub>, and half of K<sub>2</sub>SO<sub>4</sub> and heating, prior to the addition of CuSO<sub>4</sub>.5H<sub>2</sub>O and of the remaining K<sub>2</sub>SO<sub>4</sub>). In order to do this using the Buchi equipment, steps 1 and 2 of the digestion method (Table 2) were performed without the suction module of the SpeedDigester, and then CuSO<sub>4</sub>.5H<sub>2</sub>O and the remaining K<sub>2</sub>SO<sub>4</sub> were added and the digestion proceeded according to step 3 (Table 2) using the suction module. Since no recovery improvements were obtained, the method was kept as shown in Table 2, with the addition of all reagents, connection of the Buchi suction module, and start of the Buchi method.

#### Raney catalyst macromethod: catalysts

According to the MAPA manual, the macromethod requires sample masses up to 2.0 g, as well as 1.7 g of Raney catalyst and 150 mL of H<sub>2</sub>SO<sub>4</sub>-K<sub>2</sub>SO<sub>4</sub> solution (equivalent to 15 mL H<sub>2</sub>SO<sub>4</sub> and 8.0025 g K<sub>2</sub>SO<sub>4</sub>). Additionally, 1.0 g CuSO<sub>4</sub>.5H<sub>2</sub>O and 15 g K<sub>2</sub>SO<sub>4</sub> are also required in case of granular fertilizers (Chapter I of the Manual), while only 15 g K<sub>2</sub>SO<sub>4</sub> (without copper) are added in case of liquid fertilizers (chapter II of the Manual). For nitrate containing fertilizers, it is essential to keep a proportion of 1.7 g (or at least 1.5 g) of the Raney catalyst to 42 mg nitrogen (BRABSON & WOODIS, 1969; JOHNSON et al., 1974). In the Buchi SpeedDigester, it was more convenient to use half of the masses and volumes described in the MAPA manual, in order to ensure that the volume inside the digestion tube is less than 1/3 of its volume, as recommended in the Buchi manual.

It was observed that  $K_2SO_4$  and  $CuSO_4.5H_2O$  (whenever used) can be substituted by one Merck Kjeldahl tablet. These tablets contain 47.7%  $Na_2SO_4$ , 47.7%  $K_2SO_4$ , 2.8% TiO<sub>2</sub> and 1.8% CuSO<sub>4</sub>. Since each tablet weights approximately 5 g, it corresponds to 2.385 g  $Na_2SO_4$ , 2.385 g  $K_2SO_4$ , 0.14 g TiO<sub>2</sub>, and 0.09 CuSO<sub>4</sub> (equivalent to 0.1407 g CuSO<sub>4</sub>.5H<sub>2</sub>O). Although the addition of CuSO<sub>4</sub> to the analysis of liquid fertilizers strongly reduces the recovery of nitrogen, the use of the Merck Kjeldahl tablet containing CuSO<sub>4</sub> did not affect significantly the nitrogen recovery in liquid fertilizers. The tablet also contains TiO<sub>2</sub>, and the combination of CuSO<sub>4</sub> and TiO<sub>2</sub> was already reported to make protein digestion feasible in 40 minutes instead of 90 minutes in case only CuSO<sub>4</sub> was used (KANE, 1986).

### Raney catalyst macromethod: distillation

After digestion and cool down, water and NaOH are added, in order to neutralize the acid and generate an excess of NaOH (basic conditions). The addition of thiosulfate is based on its capacity to precipitate mercury, catalyst in the original method (BRABSON & WOODIS, 1969), and to allow the liberation of ammonia by boiling (HILLER et al., 1948). We tested the use of 3 mL and 12.5 mL of a 8 g/L solution of Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub>.5H<sub>2</sub>O for the macromethod and, since no recovery differences were observed, the volume of 3 mL was maintained (Table 3).

### Nitrogen recoveries

Table 5 shows the nitrogen recoveries obtained using the optimized experimental conditions summarized in Tables 2, 3 and 4. The KNO<sub>3</sub> recovery using the micromethod was the reference used during the optimization of the macromethod for KNO<sub>3</sub> and NIST 193.

Experimental conditions								Nitrogen content					
Method	N type	Sample uptake	Raney catalyst (g)	H2SO4 K2SO4 sol. (mL)	K <sub>2</sub> SO4 (g)	CuSO4. 5H2O (g)	Declared amount (% w/w)	Obtained amount (% w/w)	<i>f</i> rес (%)	Obtained amount (% w/w) average	frec (%) average	CV (%)	
Micro method	KNO3	1 g in 250 mL, aliquot 25 mL	0.7	-	-	-	13.72           13.72           13.72           13.72           13.72           13.72           13.72           13.72           13.72           13.72           13.72           13.72           13.72           13.72           13.72           13.72           13.72	12.37 12.72 13.40 13.22 13.46 13.11 14.10 14.72	90.16 92.71 97.67 96.36 98.10 95.55 102.77 107.29	13.38	97.50	5.21	

Table 5. Nitrogen recoveries.

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		Exper	imental co	nditions	Nitrogen content							
Method	N type	Sample uptake	Raney catalyst (g)	H2SO4 K2SO4 sol. (mL)	K <sub>2</sub> SO4 (g)	CuSO4. 5H2O (g)	Declared amount (% w/w)	Obtained amount (% w/w)	frec (%)	Obtained amount (% w/w) average	frec (%) average	CV (%)
							13.72	13.29	96.87			
						<u> </u>	11.40	11.98	105.06			
Macro							11.40	12.04	105.58			
method	MAP	0.5 g	0.85	75	7.5	0.5	11.40	11.29	99.01	11.77	103.19	2.89
							11.40	11.76	103.13			
							46.30	44.50	96.12			
							46.30	44.49	96.09		96.37	0.23
Macro method				75	7.5	0.5	46.30	44.76	96.68			
	Urea	0.5 g	0.85				46.30	44.67	96.48	44.62		
							46.30	44.64	96.42			
							46.30	44.64	96.42			
							20.75	20.76	100.04			
							20.75	20.72	99.85			
Macro	N-	0.5 g	0.85	75	7.5	0.5	20.75	20.88	100.62	20.80	100.24	0.30
method	$\mathrm{NH}_{4^+}$	U					20.75	20.82	100.33			
							20.75	20.83	100.38			
						<u> </u>	13.72	13.30	96.94			
							13.72	12.38	90.23			
							13.72	13.27	96.72			
							13.72	13.29	96.87			1.74
							13.72	13.28	96.79			
							13.72	12.94	94.31			
							13.72	13.10	95.48			
		0.15 g					13.72	13.31	97.01			
Macro		KNO <sub>3</sub>					13.72	13.15	95.85			
method	KNO <sub>3</sub>	(21 mg	0.85	75	7.5	0.5	13.72	12.98	94.61	13.12	95.64	
memou		(21 mg N)					13.72	13.16	95.92			
		)					13.72	13.25	96.57			
							13.72	13.25	96.72			
							13.72	13.11	90.72 95.55			
							13.72	13.02	94.90			
							13.72	13.30	94.90 96.94			
							13.72	12.89	90.94 93.95			
							13.72	12.89	95.95 96.14			
							15.72	13.19	90.14			

#### TOTAL NITROGEN DETERMINATION IN AMMONIACAL, AMIDIC AND NITRATE NITROGEN CONTAINING FERTILIZERS BY THE RANEY CATALYST METHOD

		Exper	rimental co	nditions		N	litrogen o	content				
Method	N type	Sample uptake	Raney catalyst (g)	H2SO4 K2SO4 sol. (mL)	K <sub>2</sub> SO4 (g)	CuSO4. 5H2O (g)	Declared amount (% w/w)	Obtained amount (% w/w)	<i>f</i> rec (%)	Obtained amount (% w/w) average	frec (%) average	CV (%)
							13.84	13.22	95.52	-		
							13.84	13.47	97.33			
							13.84	12.46	90.03			
							13.84	13.31	96.17			
							13.84	13.08	94.51			
							13.84	12.78	92.34			
							13.84	12.91	93.28			
	NIST	0.15 g					13.84	13.40	96.82			
Macro	193	KNO <sub>3</sub>	0.85	75	7.5	0.5	13.84	13.13	94.87	13.14	94.97	2.07
method	(N-	(N- (21 mg	15	7.5	0.5	13.84	13.19	95.30	15.14	)7.)7	2.07	
	NO3 <sup>-</sup> )	N)					13.84	13.15	95.01			
							13.84	13.16	95.09			
							13.84	12.85	92.85			
							13.84	13.66	98.70			
							13.84	13.35	96.46			
							13.84	13.16	95.09			
							13.84	13.18	95.23			
							13.84	13.14	94.94			

### CONCLUSION

The determination of nitrogen in fertilizers containing nitrogen nitrate (N-NO<sub>3</sub><sup>-</sup>) demands some attention concerning the digestion temperature, sample uptake, and exposure to humidity. The Raney macromethod was optimized for the main fertilizers nitrogen types (ammoniacal, amidic, and nitrate) using Buchi equipment in order to reach similar recoveries obtained using the Raney catalyst micromethod.

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### REFERENCES

- ABRAMS, D., METCALF, D.; HOJJATIE, M. 2014. Determination of Kjeldahl nitrogen in fertilizers by AOAC official method SM 978.02: Effect of copper sulfate as a catalyst. **Journal of the AOAC International**, Oxford, v.97, p.764-767.
- BABRAUSKAS, V. 2016. Explosions of ammonium nitrate fertilizer in storage or transportation are preventable accidents. Journal of Hazardours Materials, Amsterdam, v.304, p.134-149.
- BARKER, A. V.; VOLK, R. J. 1964. Determination of ammonium, amide, amino, and nitrate nitrogen in plant extracts by a modified Kjeldahl method. **Analytical Chemistry**, Washington D. C., v.36, n.2, p.439-439.
- BRABSON, J. A.; BURCH, W. G. Jr. 1964. Reduction of nitrates in acid medium with Raney catalyst powders. Journal of the Association of the Official Analytical Chemists, Oxford, v.47, n.6, p.1035-1040.
- BRABSON, J. A.; WOODIS Jr, T. C. 1969. A new approach to the Raney catalyst powder method for total nitrogen in fertilizers. Journal of the Association of the Official Analytical Chemists, Oxford, v.52, n.1, p.23-30.
- CAMPBELL, W. R.; HANNA, M. I. 1937. The determination of nitrogen by modified Kjeldahal methods. Journal of Biological Chemistry, Rockville, v.119, p.1-7.
- DICKINSON, W. E. 1954. Kjeldahl method as applied to the determination of nitrogen in nitrates. **Analytical Chemistry**, Washington D. C., v.26, n.4, p.777-779.
- HILLER, A.; PLAZIN, J.; VAN SLYKE, D. D. 1948. A study of conditions for Kjeldahl determination of nitrogen in proteins; description of methods with mercury as catalyst, and titrimetric and gasometric measurements of the ammonia formed. **Journal of Biological Chemistry**, Rockville, v.176, p.1401-1420.
- JOHNSON, F. J.; MILLER, D. L. 1974. Alkaline distillation of ammoniacal and nitrate nitrogen with Raney catalyst powder. **Journal of the Association of the Official Analytical Chemists**, Oxford, v.57, n.1, p.8-9.
- JOHNSON, F. J.; WOODIS, T. C.; CUMMINGS Jr., J. M. 1974. Mercury-free Raney catalyst method for total nitrogen in fertilizers. Journal of the Association of the Official Analytical Chemists, Oxford, v.57, n.1, p.10-13.
- KANE, P. F. 1986. CuSO<sub>4</sub>-TiO<sub>2</sub> as Kjeldahl digestion catalyst in manual determination of crude protein in animal feeds. Journal of the Association of the Official Analytical Chemists, Oxford, v.69, n.4, p.664-666.

- KIRK, P. L. 1950. Kjeldahl method for total nitrogen. **Analytical Chemistry**, Washington D. C., v.22, n.2, p.354-358.
- McKENZIE, H. A. 1994. The Kjeldahl determination of nitrogen: Retrospect and prospect. **Trends in Analytical Chemistry**, Amsterdam, v.13, n.4, p.138-144.
- MINISTÉRIO DA AGRICULTURA, PECUÁRIA E ABASTECIMENTO (MAPA). Secretaria de Defesa Agropecuária (SDA). Manual de Métodos Analíticos Oficiais para Fertilizantes e Corretivos. Instrução normativa Nº. 13, de 15 de julho de 2015. Brasília: **MAPA**, 2017.
- MINISTÉRIO DA AGRICULTURA, PECUÁRIA E ABASTECIMENTO (MAPA). Secretaria de Defesa Agropecuária (SDA). Instrução normativa Nº. 37, de 13 de outubro de 2017. Brasília: **Diário Oficial da União**, 13 de outubro de 2017.
- RANKER, E. R. 1926. Determination of total nitrogen, nitrate-nitrogen, and total nitrogen not including nitrate-nitrogen: further observations on a modification of the official salicylic-thiosulphate method. **Annals of the Missouri Botanical Garden**, St. Louis, v.13, n.4, p.391-424.
- SÁEZ-PLAZA, P.; MICHALOWSKI, T.; NAVAS, M. J.; ASUERO, A. G.; WYBRANIEC, S. 2013. An overview of the Kjeldahl method of nitrogen determination. Part I. Early history, chemistry of the procedure, and titrimetric finish. Critical Reviews in Analytical Chemistry, Abindon-on-Thames, v.43, n.4, p.178-223.
- SAH, R. N. 1994. Nitrate-nitrogen determination: a critical review. Communications in Soil Science and Plant Analysis, Abindon-on-Thames, v.25, p.2841-2869.

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